



Hematological Parameters and Biochemical Evaluation of the Effects of Vitamin C in Swiss Albino Mice Exposed to Chronic Doses of Cadmium

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Abstract: Cadmium toxicity characterizes a global environmental contamination delinquent and a common source of industrial and non-occupational neurological diseases. Cadmium toxicity is common among heavy metal toxicity even at minimum exposure because of slow excretion rate from the body and long biological half-life. The purpose of the present study was to determine the role of vitamin C against cadmium-induced toxicity in Swiss albino mice. This current study was performed by the biochemical parameters and hematological analysis in Swiss albino mice as affected by the oral administration of a single dose equivalent to 1/20 from LD₅₀ (100 mg/kg B.W.) of Cadmium individually for 35 days and evaluated the protective role of vitamin C. The results presented a significant reduction ($P < 0.05$) in White Blood Cells count (W.B.C.s) and Red Blood Cell (R.B.C.s) count in Cadmium-treated animals in comparison with the control group. The results from other studies showed a significant reduction of the body weight of cadmium treated mice and were compared with the control group. While total serum protein and total cholesterol significantly decreased ($P < 0.05$) in cadmium-treated animals compared with the control group. Biochemical parameters showed a significant increase ($P < 0.05$) in urea and creatinine levels in cadmium treated mice when compared to the control group. Vitamin C is recognized an essential nutrients for all species of animals. In other words, these vitamins have been shown to have protective effect against heavy metal induced toxicity. In conclusion, this study demonstrates that oral exposure of cadmium caused reduction in biochemical and hematology activities in mice and vitamin C has ameliorative effect against metal-induced toxicity. Vitamin C is a natural antioxidant that prevents the increased production of free radicals induced by oxidative damage to lipid and lipoproteins in numerous tissues and cellular compartments.

Keywords: Cadmium, Toxicity, Protective, Vitamin C, Antioxidant agent.

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1. INTRODUCTION

Environmental pollution, specifically by chemicals, is one of the most significant operative factors in destroying the biosphere mechanisms¹. Among all substances polluted, heavy metals are considered as potential unsafe contaminants in the environment to human health². Heavy metals are metallic components with density at least five times more than water³⁻⁴. In general, heavy metals are not biodegradable, have long biological half-life, and can accumulate in the different vital organs and results in unwanted side effects⁵⁻⁷. The numerous harmful health effects upon exposure to toxic heavy metals in the environment are serious concerns⁸. Much importance has been given to elucidate the toxicity mechanism due to collective environmental toxicants and mature a harmless chemotherapeutic method to mitigate the toxic belongings⁸. Cadmium is the most abundant toxic metal in the environment⁹. The anthropogenic activities and vehicular emissions contribute to poisonous metals to humans and other animal food chains¹⁰. Lead, cadmium, nickel, arsenic, chromium and mercury are the most prevalent metals that can threaten the human at low concentration¹¹⁻¹⁴. Cadmium is a significant environmental pollutant present in the soil, water, air and food¹⁵⁻¹⁶. The primary source of toxic cadmium exposure is the inhalation route of cadmium particles or fumes during industrial operations¹⁷⁻¹⁸. It is also at present in cigarette smoke¹⁹. Cadmium-induced tissue has been capable in portion to toxicant-induced oxidative stress²⁰⁻²¹. Cadmium (Cd) stimulates the development of reactive oxygen species (ROS) and metallothionein, thus causing oxidative damage to erythrocytes and various tissues consequential to the membrane functions²². Long-term exposure to Cd rises lipid peroxidation and causes inhibition of SOD (superoxide dismutase) activity, representative oxidative damage in the testes, kidney, and liver²³. Many studies suggested that the generation of reactive oxygen species (ROS) and its interference with the cellular antioxidant system is one central mechanism by which cadmium's toxic effect is mediated²⁴. The antioxidant can play a significant role in the treatment of metal-induced oxidative stress as efficient chelators²⁵. Ascorbic acid is a water-soluble dietary antioxidant that plays an essential role in controlling oxidative stress²⁶. Ascorbic acid is a necessary constituent in cellular metabolism; biomolecules' interactions give a good idea of toxicant stress and its effect²⁵. It has also been demonstrated that vitamin C is one of the greatest influential factors reducing a greater renal and hepatic cadmium problem in pigs fed diet developed with copper²⁷. Ascorbic acid is a vital phytonutrient for living cells' metabolism in diverse absorptions in natural foods, mostly fruits and their foodstuffs. Vitamin C or ascorbic acid presents a shielding consequence against free radical-induced oxidative damage²⁸. Vitamin C acts as a potent water-soluble antioxidant by scavenging reactive oxygen and nitrogen species²⁹. It is an excellent source of electrons and thus provides an electron to free radicals such as hydroxyl radical and superoxide radical and quenches their awareness³⁰. Vitamin C offers adequate protection against lipid peroxidation³¹. In addition to scavenging action, vitamin C can regenerate other small molecule antioxidants such as α -tocopherol, glutathione and urate from their respective radical species²⁹. Vitamin C is an important dietary antioxidant and significantly reduces the effect of reactive oxygen species that can cause oxidative damage such as DNA, protein and lipids which are implicated in various disease and reduces toxic effects of metal³²⁻³³. This study aimed to investigate a possible protective effect of

Vitamin C treatment on the selected biochemical parameters and histological changes of RBCs and WBCs in Swiss albino mice exposed to Cd.

2. MATERIALS AND METHOD

2.1 Chemical

Cadmium, Vitamin C (Ascorbic acid), and all the chemicals used in the experiments were of the analytical grade and purchased from Himedia Laboratories Private Limited. (Mumbai, India).

2.2 Animals

Swiss albino mice (5-6 weeks old, weighing almost 20-25 gm) were housed in polypropylene cages (3-4 animals per cage; B.I.K Industries). The animals were kept throughout the experiment for full accommodation in an air-conditioned animal room ($28\pm 2^\circ\text{C}$)³⁴ and controlled light room with a photoperiod of under a 12 h light/dark cycle and given a standard pellet diet (obtained from Hindustan Lever Limited, Mumbai, India) and RO water *ad libitum*. All experimental processes were completed following the recommendations found in the Guide for the Care and Use of Laboratory Animals (Refer) and permitted by committee Institutional Animal Ethics Committee (IAEC) (R. No. 1402/a/10/CPCSEA) of the Jayoti Vidyapeeth Women's University of Jaipur. Established ethical strategies were also followed in all Experiments.

2.3 Experimental design

The Swiss albino mice were divided into three groups comprising of 4 animals in each group as follow:-

Group I: - Control (normal) swiss albino mice, fed with standard pellet diet and water.

Group II: - Were given 100mg/kg body weight of Cadmium chloride.

Group III:- was given 100mg/kg body weight of the Vitamin C after exposure to Cadmium.

2.4 Blood Sample Collection

The mice were carefully monitored every day. Animals defined as fasted were destitute from food for at least 12 h but an acceptable free entree to drinking water. Blood samples were collected at weekly intermissions till the end of the research work³⁴. During the 7th, 14th, 28 and 35 week of treatment, the body weight, Serum Total Cholesterol, Total serum protein, Total blood urea, and serum creatinine of all the Swiss albino mice were determined. The blood samples were poised from the tail vein puncture. Throughout the experiment, blood samples were been obtained overnight from the tail vein of all the animals. Blood was left to clot and centrifuged (REMI CM12;2009) at 3000 rpm for 15 min, at 4°C for separating the serum, frozen and stored at -20°C ³⁴. for biochemical analysis and hematological analysis.

2.5 Bodyweight Estimation

All animals were weighed every week until the end of the experiment.

2.6 Biochemical Estimation

The blood sample was collected for Hematological Parameters: red blood cells count (RBCs), and white blood cell count (WBCs) was determined according to the methods described by previous research studies³⁵. Serum was collected for estimation of biochemical parameters, such

as serum cholesterol³⁶, total serum protein³⁷; serum creatinine³⁸ and blood urea level³⁹.

3 STATISTICAL ANALYSIS

The results were expressed as mean ± SEM (Standard Error of Mean). The data were analyzed by one-way ANOVA followed by student t-test at the level of significance was described as P<0.05 and P<0.01. All statistical data were performed using IBM SPSS Statistics 20 (File version 22.0.0.0).

4 RESULT

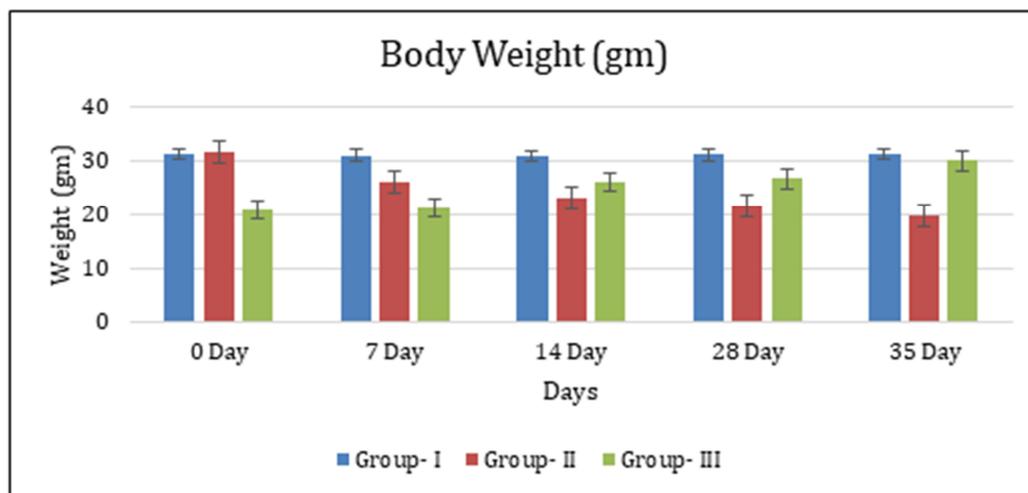
In this investigation, the bodyweight of albino mice subjected to treatment is shown in Table I and Graph I. In Cd-treated

Animals, the results showed a significant decrease (p < 0.05) in body weight compared to the control group. However, vitamin C supplements the body weight gain significantly (p < 0.05) than in albino mice exposed to cadmium p < 0.05. In the present study, the Red Blood cell (RBC) and White Blood cell (WBC) of Swiss albino mice subjected to Cadmium treatments are shown in Table 2 and Figure 2. During the experiment, the White Blood cell (WBC) and Red Blood cell (RBC) of the Swiss albino mice group (9.06±0.001; 8.08±0.004) slightly significantly decreased as compared with the control group (3.79±0.001; 3.53±0.009) (p<0.05). Vit C treatment increased (p<0.05) the lowered RBC counts. It was also found that the WBC counts increased (p<0.05) in Cd-treated rats.

Table I: The effect of Vitamin C on Body Weight (gm) in Cadmium treated Mice.

Days	Group- I Control	Group- II Cadmium treated Mice	Group- III Vitamin C treated Mice
0 Day	31.18±0.09	31.56±0.09	20.79±0.02
7 Day	31.09±0.07	26.17±0.05	21.18±0.04
14 Day	30.81±0.06	23.19±0.04	25.97±0.05
28 Day	31.13±0.07	21.62±0.04	26.67±0.05*
35 Day	31.25±0.09	19.79±0.01	29.98±0.06*

Values (In Gram) are shown as the Mean±SEM. Values are statistically significant at * p<0.05 when compared to control group.

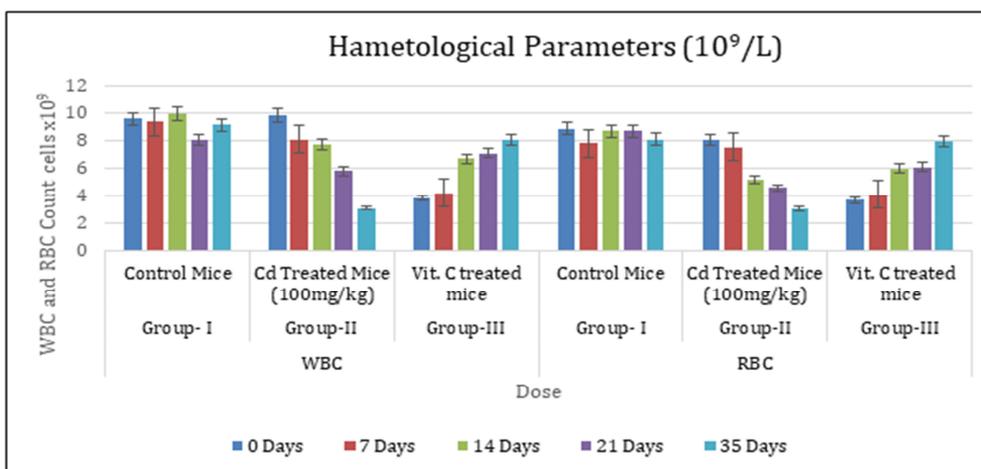


Graph I: Showing the effect of Vitamin C on Body weight in Cadmium treated mice

Table 2: The effect of Vitamin C on Hematological Parameters in Cadmium treated Mice.

Days	WBC			RBC		
	Group- I	Group-II	Group-III	Group- I	Group-II	Group-III
	Normal Mice	Cd Treated Mice (100mg/kg)	Vitamin C treated mice	Control Mice	Cd Treated Mice (100mg/kg)	Vitamin C treated mice
0 Days	9.57±0.009	9.85±0.009	3.78±0.003	8.88±0.007	8.02±0.007	3.64±0.003
7 Days	9.36±0.008	8.09±0.007	4.18±0.004	7.78±0.006	7.53±0.005	4.07±0.005
14 Days	9.95±0.011	7.75±0.006	6.67±0.005	8.68±0.008	5.16±0.004	5.98±0.006
21 Days	8.08±0.007	5.76±0.004	7.08±0.006*	8.67±0.008	4.54±0.004	6.08±0.006*
35 Days	9.13±0.008	3.07±0.003	8.02±0.007*	8.09±0.007	3.02±0.002	7.98±0.008*

Values are shown as the Mean±SEM. Values are statistically significant at * p<0.05 when compared to the Normal group.



Graph 2: Effect of Vitamin C on Hematological parameters in Cadmium treated mice

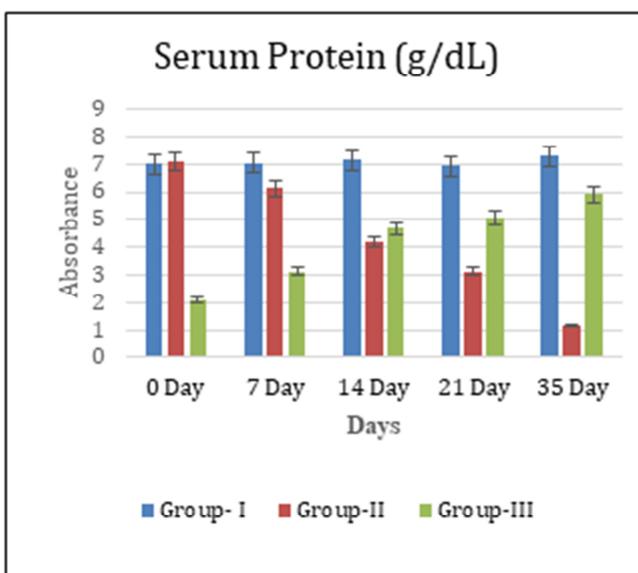
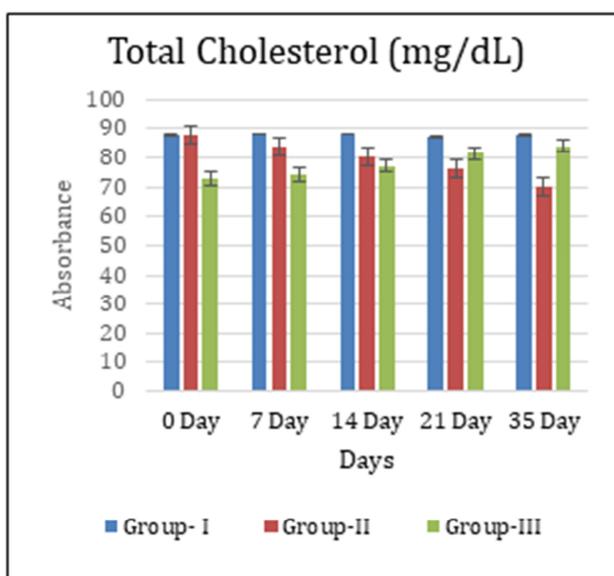
Treatment with Cd caused a significant decrease of cholesterol (70.09±0.11) when (p<0.05) compared to the control group (87.09±0.24). Meanwhile, the concentration of total serum protein was reduced (2.11±0.001) (p < 0.05) as compared to the control group (7.13±0.009). The administration of vitamin C in cadmium-treated mice increased cholesterol level and the serum protein levels were in compared to cadmium treated mice. The results showed the serum creatinine Cd-treated mice group significantly increased (6.76±0.21) concerning the control group

(0.98±0.08) (P<0.05). However, vitamin C supplies the serum creatinine became significantly decrease (p < 0.05) than in albino mice exposed to cadmium p < 0.05. Cadmium induced mice were found to have significantly elevated blood urea level (57.09±0.28) with the respect to the control group (38.99±0.13) (p<0.05) Table 3; Figure 3. The administration of vitamin C in cadmium treated albino mice increased blood urea level was significantly decreased as compared to normal mice. (p<0.05) Table 4; Figure 4.

Table 3: The effect of Vitamin C on Total Cholesterol and Total Serum Protein in Cadmium treated Mice.

Days	Total Cholesterol (mg/dL)			Total Serum Protein (g/dL)		
	Group- I	Group-II	Group-III	Group- I	Group-II	Group-III
	Control	Cadmium treated Mice	Vit. C treated mice	Control	Cadmium treated Mice	Vit. C treated mice
0 Day	87.92±0.17	88.02±0.18	72.78±0.11	6.98±0.007	7.09±0.008	2.11±0.001
7 Day	88.34±0.19	84.05±0.14	74.12±0.12	7.03±0.009	6.11±0.008	3.12±0.003
14 Day	88.08±0.18	80.15±0.13	77.08±0.12	7.13±0.009	4.19±0.005	4.68±0.004
21 Day	87.16±0.17	76.16±0.11	81.56±0.14*	6.92±0.008	3.12±0.003	5.08±0.004*
35 Day	87.68±0.17	70.09±0.10	84.11±0.15*	7.29±0.009	1.14±0.001	5.91±0.005*

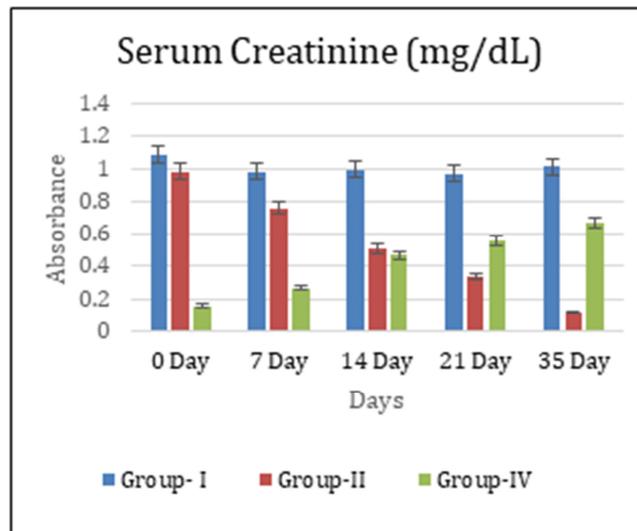
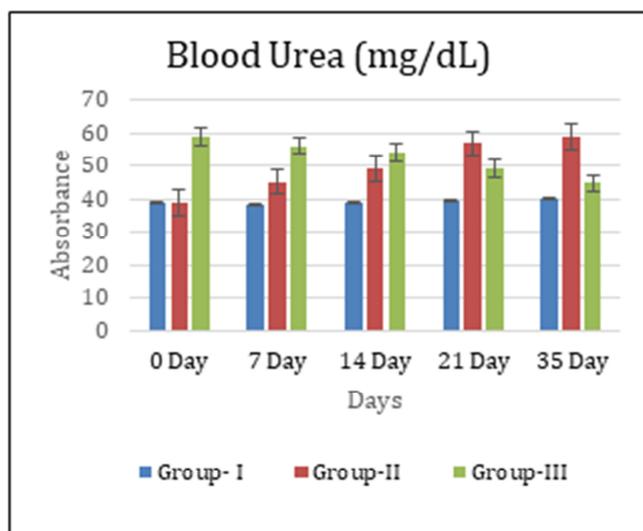
Values are shown as the Mean±SEM. Values are statistically significant at * p<0.05 when compared to the normal group.



Graph 3: Effect of Vitamin C on Total Cholesterol and Total Serum Protein in normal mice, Cadmium treated mice and Vitamin treated mice

Table 4: The effect of Vitamin C on Serum Blood Urea and Serum Creatinine in Cadmium treated Mice.

Days	Serum Blood Urea (mg/dL)			Serum Creatinine mg/dL		
	Group- I	Group-II	Group-III	Group- I	Group-II	Group-IV
	Control	Cadmium treated Mice	Vit. C treated mice	Control	Cadmium treated Mice	Vit. C treated mice
0 Day	39.08±0.11	38.97±0.09	58.98±0.19	1.09±0.006	0.98±0.006	0.16±0.001
7 Day	38.19±0.09	45.09±0.14	56.03±0.17	0.98±0.005	0.76±0.005	0.27±0.002
14 Day	38.91±0.09	49.17±0.15	54.01±0.16	0.99±0.005	0.51±0.005	0.47±0.003
21 Day	39.78±0.10	56.98±0.18	49.18±0.15*	0.97±0.004	0.34±0.003	0.56±0.004*
35 Day	40.01±0.12	59.01±0.19	44.77±0.13*	1.01±0.007	0.12±0.001	0.67±0.005*



Graph 4: Effect of Vitamin C on Serum Blood Urea and Serum Creatinine in normal mice, Cadmium treated mice and Vitamin treated mice

5 DISCUSSION

Different people have clarified the mechanism of cadmium-induced hepatotoxicity which includes contact with membrane components and lipid peroxidation⁴⁰. Cadmium induces oxidative stress and lipid peroxidation by reducing GSH (Glutathione) or by inhibition of antioxidant enzymes⁴⁰. Cadmium becomes accrued in liver and causes tissue damage⁴¹. Cadmium indirectly generates various radicals like superoxide, hydroxyl and nitric oxide and induces oxidative stress and tissue damage⁴². These reactive oxygen species generated indirectly by Cd, attack on the cell membrane and causes destabilization and disintegration of the cell membrane resulting in lipid peroxidation^{43,16}. Because of its oxidative stress inducing nature the cadmium induced toxicity can be restored by the treatment of various antioxidant^{44,45}. The present study also confirmed that the administration of vitamin C significantly reduces the toxicities induced by cadmium. The increase in the pollution of our daytime is an important and worldwide problem. It is outstanding for using xenobiotic substances or deadly chemicals or by certain synthetic compounds such as heavy metals⁴⁶⁻⁴⁷. Cadmium and lead are very toxic heavy metals and a significant environmental pollutant that causes extermination in numerous tissues of animals and human⁴⁸⁻⁴⁹. Vitamin C is an effective water solvable antioxidant created in human plasma. It scavenges reactive oxygen and nitrogen species including hydroxyl radical, peroxy radicals, superoxide anion, nitrogen dioxide as well as non-radical species such as hypochlorous acid, ozone and singlet oxygen⁵⁰. In the current study, Cadmium fed mice were exposed to a substantial and constant decrease in body weight at different intervals over the control group's experiment period. An earlier report has

been described that in swiss albino mice, that significantly reduces body weight⁵¹⁻⁵². The reduction in body weight content in the Cd-treated mice group correlates with other author's work⁵³⁻⁵⁴. In the current work, the investigation of samples obtained from animals treated with vitamin C showed enhancement in the body weight as compared to the cadmium mice group. These results agreed with the further study, which found that the ascorbic acid prohibited the free radicals caused oxidative impairment of the cell membrane³⁹. The treatment with vitamin C to the Cadmium-treated animals enhanced body weights^{55,1}. This outcome agrees with other results⁵⁶⁻⁵⁷. Vitamin C supplementation facilitated a significant increase in body weight compared to cadmium treated mice^{58,53}. The observed rats reduced weight gain in this study is reliable with some earlier published reports⁵⁹. The present study result exposed a significant decrease in WBC and RBC of treated mice with Cadmium compared with the control mice group. It has been described that RBC and WBC count significantly reduced in the swiss albino mice administered CdCl₂^{60,51}. On the other hand, These outcomes approve with those found by early studies^{54,58}. They specified that the RBC count significantly reduced in Wistar Rats, showing the Cd concentration relative to the control mice group. Other authors revealed this outcomes⁶¹⁻⁶² conducted on mice poisoned with lead. In the current search, the Vitamin C treated group showed a rise in hematological parameters compared to cadmium-treated mice. Another work found that hematological parameters significantly increased by dietary vitamin C⁵⁴. On the other hand, it was also shown that pre-treatment with vitamin C showed a defensive role on the toxicity of Cadmium on hematological value⁶³. Similar outcomes were obtained by earlier published works⁶⁴, which exposed the protective effect of vitamin C on

anemia produced by heavy metals in rats. The previous report showed that treatment with Vitamin C showed meaningful reduction of the toxic effects of Cadmium on hematological standards⁵⁸. In Our current study cadmium treated mice presented a significant reduction in Cholesterol, serum protein, increased serum creatinine and blood urea level compared to the control mice group. In the present work, examining specimens obtained from animals treated with vitamin C showed improvement in the biochemical parameters compared to the cadmium group. A similar result has been reported a significant decrease in total serum protein level and increased urea and creatinine levels in albino rats^{65-67,1}. These results are in coincidence with those previously obtained study¹. They found that rats treated with vitamin C showed a significant increase in total serum protein level and decreased urea and creatinine levels in albino rats. The reduction in cholesterol content in the Cd-treated mice group agrees with other authors' work^{67,68-71}. Reduced protein content was observed in the Cd-treated group. Some authors also reported a similar trend in their research findings^{69-71,72-75}. This result agrees with other findings^{76,65}. On the other hand, the results obtained showed a significant increase in total serum protein level by the treatment of vitamin C on Cadmium-induced rats⁷⁷. Another worker reported that the administration of vitamin C significantly decreases urea and creatinine levels in Wistar rats⁷⁸.

6 CONCLUSION

The outcomes of this current study showed that cadmium present in the atmosphere and in the specific products are the cause of hematological and biochemical disorders in the blood. Vitamin C has a protective effect on biochemical and hematological induced cadmium toxicity. The present work also presented that Vitamin C has a significant result on the

11 REFERENCES

- Sharaf AMM, Farrag A-RH, Fahmy HM. Protective effects of vitamin C on hematological and biochemical parameters of intoxicated male albino rats with lead and cadmium. *Middle East. J Appl Sci.* 2017;7(1):57-67.
- Feleafel MN, Mirdad ZM. Hazard and Effects of Pollution by Lead on Vegetable Crops. *J Agric Environ Ethics.* 2013;26(3):547-67. doi: [10.1007/s10806-012-9403-1](https://doi.org/10.1007/s10806-012-9403-1).
- Pouls M. 'Extended health.' A website for doctors and health professionals Al/meat-10.htm. 2005;2005:1-25.
- Igwegbe AO, Agukwe CH, Negbenebor A. A survey of heavy metal (lead, cadmium and copper) contents of selected fruit and vegetables crops from Borno State of Nigeria. *Int J Eng Sci.* 2013;2(1):01-5.
- Sathawawara NG, Parikh DJ, Agarwal YK. Essential heavy metals in environmental samples from Western India. *Bull. Environ Contam Toxicol.* 2004;73:756-61.
- Bagdatlioglu N, Nergiz C, Ergonul PG. Heavy metal levels in leafy vegetables and some selected fruits. *J Verbr Lebensm.* 2010;5(3-4):421-8. doi: [10.1007/s00003-010-0594-y](https://doi.org/10.1007/s00003-010-0594-y).
- Radwan MA, Salama AK. Market basket survey for some heavy metals in Egyptian fruits and vegetables. *Food Chem Toxicol.* 2006;44(8):1273-8. doi: [10.1016/j.fct.2006.02.004](https://doi.org/10.1016/j.fct.2006.02.004), PMID [16600459](https://pubmed.ncbi.nlm.nih.gov/16600459/).
- Patra RC, Rautray AK, Swarup D. Oxidative stress in lead and cadmium toxicity and its amelioration. *Vet Med Int.* 2011;20:457327. doi: [10.4061/2011/457327](https://doi.org/10.4061/2011/457327). PMID: [21547215](https://pubmed.ncbi.nlm.nih.gov/21547215/).
- Masoomah Masoomi Karimi, Moslem Jafari Sani, Ali Zaree Mahmudabadi, Asma Jafari sani, Seyed Reza Khatibi. Effect of Acute Toxicity of Cadmium in Mice Kidney Cells. *Iranian Journal of Toxicology.* 2012; 6(18).
- Okada IA, Sakuma AM, Maid FD, Dovidemskas S, Zenebon O. Evaluation of lead and cadmium in milk due to environmental contamination in Paraiba valley region of South Estern Brazil. *Raissade-Saúde-Publ.* 1997;31:140-3.
- Kechrid Z, Dahdouh F, Djabar RM, Bouzerna N. Combined effect of water contamination with cobalt and nickel on metabolism of albino (Wistar) rats. *Iran J Environ Health Sci Eng.* 2006;3(1):65-9.
- Das KK, Das SN, Dhundasi SA. Nickel, its adverse health effects & oxidative stress. *Indian J Med Res.* 2008;128(4):412-25. PMID [19106437](https://pubmed.ncbi.nlm.nih.gov/19106437/).
- Sattar MU, Khan MA, Khalil AA, Amir RM. Mitigation of heavy metals in vegetables through washing with house hold chemicals. *Int J Agric Sci.* 2013;3(5):1-12.
- Al-fatlawi, A.C. and M.H. Al-Murshedi. The effects of heavy metal (Nickel) on hematological parameters of laboratory male mice. *Int J Adv Res.* 2015;3(9):598-601.
- Cinar, M. Cadmium and effects at biological system. *Veterinarium.*2003;14: 79-84.
- O. Kaplan, N.C. Yildirim, N. Yildirim and M. Cimen. Toxic Elements in Animal Products and Environmental

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8 AUTHORS CONTRIBUTION STATEMENT

Both the authors have contributed equally.

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10 CONFLICT OF INTEREST

Conflict of interest declared none.

- Health. *Asian Journal of Animal and Veterinary Advances*. 2011;6: 228-232. DOI: 10.3923/ajava.2011.228.232.
17. Office of Environmental Health Hazard Assessment (OEHA). Public health goal for cadmium in drinking water; Feb 1999.
 18. Mckenna IM, Waalkes MP, Chen LC, Gordon T. Comparison of inflammatory lung responses in Wistar rats and C57 and DBA mice following acute exposure to cadmium oxide fumes. *Toxicol Appl Pharmacol*. 1997;146(2):196-206. doi: [10.1006/taap.1997.8241](https://doi.org/10.1006/taap.1997.8241), PMID 9344887.
 19. Stohs SJ, Bagchi D, Bagchi M. Toxicity of trace element in tobacco smoke. *Inhal Toxicol*. 1997;9(9):867-90. doi: [10.1080/089583797197926](https://doi.org/10.1080/089583797197926).
 20. Patra RC, Swarup D, Dwivedi SK. Antioxidant effects of α tocopherol, ascorbic acid and L-methionine on lead induced oxidative stress to the liver, kidney and brain in rats. *Toxicology*. 2001;162(2):81-8. doi: [10.1016/s0300-483x\(01\)00345-6](https://doi.org/10.1016/s0300-483x(01)00345-6), PMID 11337108.
 21. Fu H, Ye XB, Zhu JL et al. Oxidative stress in lead exposed workers. In: IARC Gargnano Conference. Vol. 2; 1999. p. 3.
 22. Sarkar S, Yadav P, Bhatnagar D. Lipid peroxidative damage on cadmium exposure and alterations in antioxidant system in rat erythrocytes: a study with relation to time. *BioMetals*. 1998;11(2):153-7. doi: [10.1023/a:1009286130324](https://doi.org/10.1023/a:1009286130324), PMID 9542068.
 23. Patra RC, Swarup D, Senapati SK. Effects of cadmium on lipid peroxides and superoxide dismutase in hepatic, renal and testicular tissue of rats. *Vet Hum Toxicol*. 1999;41(2):65-7. PMID 10192131.
 24. Sen Gupta R, Sen Gupta ES, Dhakal BK, Thakur AR, Ahnn J. Vitamin C and vitamin E protect the rat testes from cadmium-induced reactive oxygen species. *Mol Cells*. 2004;17(1):132-9. PMID 15055539.
 25. Ramdas, Borane Vijay. Protective role of Ascorbic acid on the lead chloride induced alterations in the Ascorbic Acid contents of the fresh water fish, *Channa orientalis* (Schneider). *Advances in Applied Science Research*, 2013;4(2):305-308.
 26. Panayiotidis M, Collins AR. Ex vivo assessment of lymphocyte antioxidant status using the comet assay. *Free Radic Res*. 1997; Nov 27;27(5):533-7. doi: [10.3109/10715769709065793](https://doi.org/10.3109/10715769709065793), PMID 9518069.
 27. Kapl D, Weiser H, Rambeck WA. The influence of vitamin C on cadmium retention in pigs. *Rev Med Vétérinaire*. 1994;145:291-7.
 28. Padayatty SJ, Katz A, Wang Y, Eck P, Kwon O, Lee JH, Chen S, Corpe C, Dutta A, Dutta SK, Levine M. Vitamin C as an antioxidant: evaluation of its role in disease prevention. *J Am Coll Nutr*. 2003;22(1):18-35. doi: [10.1080/07315724.2003.10719272](https://doi.org/10.1080/07315724.2003.10719272), PMID 12569111.
 29. Halliwell B. Vitamin C: antioxidant or pro-oxidant in vivo? *Free Radic Res*. 1996;25(5):439-54. doi: [10.3109/10715769609149066](https://doi.org/10.3109/10715769609149066), PMID 8902542.
 30. Bendich A. Antioxidant micronutrients and immune responses. *Ann N Y Acad Sci*. 1990;587:168-80. doi: [10.1111/j.1749-6632.1990.tb00144.x](https://doi.org/10.1111/j.1749-6632.1990.tb00144.x), PMID 2193567.
 31. Frei B Proceedings of the Society for Experimental Biology and Medicine. On the role of vitamin C and other antioxidants in athero-genesis and vascular dysfunction. 1999;222(3):196-204.
 32. You WC, Zhang L, Gail MH, Chang YS, Liu WD, Ma JL, Li JY, Jin ML, Hu YR, Yang CS, Blaser MJ, Correa P, Blot WJ, Fraumeni JF Jr, Xu GW. Gastric dysplasia and gastric cancer: Helicobacter pylori, serum vitamin C, and other risk factors. *J Natl Cancer Inst*. 2000;4:92(19):1607-12. doi: 10.1093/jnci/92.19.1607. PMID: 11018097.
 33. Gamal H. El-Sokkary , Eatemad A. Awadalla. The Protective Role of Vitamin C Against Cerebral and Pulmonary Damage Induced by Cadmium Chloride in Male Adult Albino Rat. *The Open Neuroendocrinology Journal*, 2011;4: 1-8. [DOI: 10.2174/1876528901104010001].
 34. Pramod K. Raghav and Shilpa Bhargava. Characterization of fructose diet induced diabetes mellitus in swiss albino mice. *IJPSR*, 2015; Vol. 6(5): 2140-2145. DOI: 10.13040/IJPSR.0975-8232.6(5).2140-45.
 35. Dacie SJV, Lewis SM. *Practical haematology*. 6th ed. Churchill Livingstone; 1984. p. 22-7.
 36. Zak B. Simple Rapid Micro technic for Serum Total cholesterol. *Am J Clin Pathol*. 1957;27(5):583-8. doi: [10.1093/ajcp/27.5_ts.583](https://doi.org/10.1093/ajcp/27.5_ts.583), PMID 13435243.
 37. Lowry OH, Rosebrough NJ, Farr AL, Randall RJ. Protein measurement with the Folin phenol reagent. *J Biol Chem*. 1951;193(1):265-75. doi: [10.1016/S0021-9258\(19\)52451-6](https://doi.org/10.1016/S0021-9258(19)52451-6), PMID 14907713.
 38. Brod J, Sirota JH. The renal clearance of endogenous "creatinine" in man. *J Clin Invest*. 1948;27(5):645-54. doi: [10.1172/JCI102012](https://doi.org/10.1172/JCI102012), PMID 16695585.
 39. Natelson S, Scott ML, Beffa C. A rapid method for the estimation of urea in biologic fluids. *Am J Clin Pathol*. 1951;21(3):275-81. doi: [10.1093/ajcp/21.3_ts.275](https://doi.org/10.1093/ajcp/21.3_ts.275), PMID 14818981.
 40. Bagchi, D., Bagchi, M., Hassoun, E.A. et al. Cadmium-induced excretion of urinary lipid metabolites, DNA damage, glutathione depletion, and hepatic lipid peroxidation in sprague-dawley rats. *Biol Trace Elem Res* 52, 143 (1996). <https://doi.org/10.1007/BF02789456>.
 41. Stohs SJ, Bagchi D. Oxidative mechanisms in the toxicity of metal ions. *Free Radic Biol Med*. 1995;18(2):321-36. doi: 10.1016/0891-5849(94)00159-h. PMID: 7744317.
 42. Stohs SJ, Bagchi D, Hassoun E, Bagchi M. Oxidative mechanisms in the toxicity of chromium and cadmium ions. *J Environ Pathol Toxicol Oncol*. 2001;20(2):77-88. PMID: 11394715.
 43. Skakun NP, Vysotskiĭ Ilu. Vliianie tetratsiklinovykh antibiotikov na perekisnoe okislenie lipidov [Effect of tetracycline antibiotics on lipid peroxidation]. *Antibiotiki*. 1982;27(9):684-7. Russian. PMID: 7149691.
 44. Eriyamremu, G.E., M.A. Adaikpoh and F.O. Obi. Pretreatment of Rats with α -tocopherol alter liver and kidney protein, alkaline phosphatase activity and phospholipid profile after 24 hour intoxication with cadmium. *J. Med. Sci*. 2006; 6: 615-620. DOI: 10.3923/jms.2006.615.620.
 45. Karbownik M, Gitto E, Lewinski A, Reiter RJ. Induction of lipid peroxidation in hamster organs by the carcinogen cadmium: melioration by melatonin. *Cell Biol Toxicol*. 2001;17(1):33-40. doi: 10.1023/a:1010903130693. PMID: 11504448.
 46. Chihuailaf R, Contreras P, Wittwer F. Pathogenesis of oxidative stress: consequences and evaluation in animal health. 2002;33(3):265-83.
 47. Sen Gupta R, Sen Gupta E, Dhakal BK, Thakur AR, Ahnn J. Vitamin C and vitamin E protect the rat testes

- from cadmium-induced reactive oxygen species. *Mol Cells*. 2004;17(1):132-9. PMID [15055539](#).
48. Jagadeesan G, Sankarsami Pillai S. Hepatoprotective effects of taurine against mercury induced toxicity in rats. *J Environ Biol*. 2007;28(4):753-6. PMID [18405108](#).
 49. Akinyeye AJ, Okorie TG. Heavy metal studies of industrial effluent on Alaro stream sediment, I. *Res J Biol Sci*. 2012;1(6):5-9.
 50. Flora SJS, Flora G, Saxena G. Environmental occurrence, health effects and management of lead poisoning. In: Jose SC, Jose S, editors. *Lead*. Amsterdam: Elsevier Sci; 2006. p. 158-228.
 51. Rekha DK, Tripathi Y, Raghuvver CV, Sheil ARP, Ramaswamy C, Priya K. Role of vitamin C an antioxidant in cadmium chloride induced testicular damage. *Int J Appl Biol Pharm Technol*. 2011;2(3):484-8.
 52. Kini RD, Tripathi Y, Raghuvver CV, Pai Sheil AR, Ramaswamy C, Kamath P. Role of vitamin C as an antioxidant in cadmium chloride induced testicular damage. *Int J Appl Biol Pharm Technol*. 2011; Jul-Sep;2(3):484-8.
 53. Sarkar MAR, Khan MZH, Sharmin T, Rahman SM, Ferdousi Z. Toxicological effects of cadmium chloride on Swiss albino mice. *Mus sp. (Rodentia: Muridae)*. *Int J Environ Biol*. 2013;3(1):50-6.
 54. Kumari a, Sharma S. Curcumin protection against cadmium chloride-induced biochemical alterations in lungs of swiss albino mice. *Asian J Pharm Clin Res*. 2020:103-7.
 55. Naima L, Zine K. Combined protective effect of vitamins C and E on cadmium induced oxidative liver injury in rats. *Afr J Biotechnol*. 2012;11(93):16013-20. doi: [10.5897/AJB12.2665](#).
 56. Affi OK, Embaby AS. Histological study on the protective role of ascorbic acid on cadmium induced cerebral cortical neurotoxicity in adult male albino rats. *J Microsc Ultrastruct*. 2016;4(1):36-45. doi: [10.1016/j.jmau.2015.10.001](#), PMID [30023208](#).
 57. El-Sokkary GH, Awadalla EA. The protective role of vitamin C against cerebral and pulmonary damage induced by cadmium chloride in male adult albino rat. *TONEUROEJ*. 2011;4(1):1-8. doi: [10.2174/1876528901104010001](#).
 58. Ali S, Hussain S, Khan R, Mumtaz S, Ashraf N, Andleeb S, Shakir HA, Tahir HM, Khan MKA, Ulhaq M. Renal toxicity of heavy metals (cadmium and mercury) and their amelioration with ascorbic acid in rabbits. *Environ Sci Pollut Res Int*. 2019;26(4):3909-20. doi: [10.1007/s11356-018-3819-8](#), PMID [30547340](#).
 59. Hounkpatin ASY, Etorh PA, Guedenon P, et al. Hematological evaluation of Wistar rats exposed to chronic doses of cadmium, mercury and combined cadmium and mercury. *Afr J Biotechnol*. 2013;12(23):3731-7.
 60. Horiguchi H, Sato M, Konno N, Fukushima M. Long-term cadmium exposure induces anemia in rats through hypoinduction of erythropoietin in the kidneys. *Arch Toxicol*. 1996;71(1-2):11-9. doi: [10.1007/s002040050352](#), PMID [9010580](#).
 61. Saggu S, Rehman H, Aziz AT, Alzeibr FMA, Oyouni AAA, Zidan N, Panneerselvam C, Trivedi S. *Cymbopogon Schoenanthus (Ethkher)* ameliorates cadmium induced toxicity in swiss albino mice. *Saudi J Biol Sci*. 2019;26(7):1875-81. doi: [10.1016/j.sjbs.2017.01.002](#), PMID [31762670](#).
 62. Veena S, Leena K, Arti S, Shweta L, Sharma SH. Ameliorating effect of *Coriandrum sativum* extracts on hematological and immunological variables in an animal model of lead intoxication. *J Pharm Allied Health Sci*. 2011;1:16-29.
 63. Iavicoli I, Carelli G, Stanek EJ, Castellino N, Calabrese EJ. Effects of low doses of dietary lead on red blood cell production in male and female mice. *Toxicol Lett*. 2003;137(3):193-9. doi: [10.1016/s0378-4274\(02\)00404-6](#), PMID [12523962](#).
 64. Ognjanović BI, Pavlović SZ, Maletić SD, Zikić RV, Stajin AS, Radojčić RM, Sačić ZS, Petrović VM. Protective influence of vitamin E on antioxidant defense system in the blood of rats treated with cadmium. *Physiol Res*. 2003;52(5):563-70. PMID [14535831](#).
 65. Fox MR, Fry BE, Harland BF, Schertel ME, Weeks CE. Effect of ascorbic acid on cadmium toxicity in the young coturnix. *J Nutr*. 1971;101(10):1295-305. doi: [10.1093/jn/101.10.1295](#), PMID [5098870](#).
 66. Das KK, Dasgupta S. Effect of nickel on testicular nucleic acid concentrations of rats on protein restriction. *Biol Trace Elem Res*. 2000;73(2):175-80. doi: [10.1385/BTER:73:2:175](#), PMID [11049209](#).
 67. Dostal LA, Hopfer SM, Lin SM, Sunderman FW. Effects of nickel chloride on lactating rats and their suckling pups, and the transfer of nickel through rat milk. *Toxicol Appl Pharmacol*. 1989;101(2):220-31. doi: [10.1016/0041-008x\(89\)90271-8](#), PMID [2479122](#).
 68. Kshirsagar M, Patil J, Patil A, Ghanwat G, Sontakke A, Ayachit R. Biochemical effects of lead exposure and toxicity on battery manufacturing workers of Western Maharashtra (India): with respect to liver and kidney function tests. *Al Ameen J Med Sci*. 2015;8(2):107-14.
 69. Purohit RK, Chakrawarti A, Bhartiya KM. Radiation and cadmium induced biochemical alterations in mouse kidney. *Iran J Radiat Res*. 2007;5:125-30.
 70. Sharma S, Vijaya P. Nephrotoxic effects of cadmium. *Trends Life Sci*. 2015;4:352-7.
 71. Sharma S, Vijaya P. Ameliorating potential of lycopene against cadmium toxicity of kidney of albino mice. *Int J Adv Res*. 2015;3:766-70.
 72. Chakrawarti A, Purohit RK, Agarwal M, Joshi P, Basu A, Bhartiya KM. Modulation of radiation and cadmium induced biochemical changes in mouse kidney by *Emblca officinales* Linn, Iran. *J Radiat Res*. 2010;8:3-10.
 73. El-Demerdash FM, Yousef MI, Kedwany FS, Baghdadi HH. Cadmium-induced changes in lipid peroxidation, blood hematology, biochemical parameters and semen quality of male rats: protective role of vitamin E and beta-carotene. *Food Chem Toxicol*. 2004;42(10):1563-71. doi: [10.1016/j.fct.2004.05.001](#), PMID [15304303](#).
 74. Kaoud HA, Kamel MM, Abdel-Razek AH, Kamel GM, Ahmed KA. Neurobehavioral, neurochemical and neuromorphological effects of cadmium in male rats. *J Am Sci*. 2010;6:189-02.
 75. Somade PM, Adnaik RS, Mohite SK, Magdum CS. Protective role of *Cucumis melo* against cadmium induced oxidative neurotoxicity in mice. *Int J Univ Pharm Biosci*. 2014;3:269-79.
 76. Babaknejad N, Moshtaghie AA, Nayeri H, Hani M, Bahrami S. Protective role of zinc and magnesium against cadmium nephrotoxicity in male Wistar rats.

- Biol Trace Elem Res. 2016;174(1):112-20. doi: 10.1007/s12011-016-0671-x, PMID 27038621.
77. Yousuf MB. Effect of high dietary intake of nickel in the West African Dwarf goat. Ghana J Agric Sci. 2002;35:147-51.
78. Obianime AW, Roberts II. Antioxidants, cadmium-induced toxicity, serum biochemical and the histological abnormalities of the kidney and testes of the male Wistar rats. Niger J Physiol Sci. 2009 Dec;24(2):177-85. doi: 10.4314/njps.v24i2.52910, PMID 20234761.